A Mendelian Randomization Study Identifies the Causal Association between Plasma Mitochondrial CHCHD Proteins and Polycystic Ovary Syndrome

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ABSTRACT

Purpose: The objective of this research was to examine the causal link between Polycystic Ovary Syndrome (PCOS) and plasma mitochondrial Coiled-Coil-Helix-Coiled-Coil-Helix Domain (CHCHD) proteins using a Mendelian Randomization (MR) method.

Methods: We performed a two-sample MR analyses by utilizing summary statistics obtained from Genome-Wide Association Studies (GWAS) of PCOS (642 cases and 118,228 controls) and the levels of CHCHD2 and CHCHD10 in plasma (3,301 individuals). The Inverse-Variance Weighted (IVW) method was used for the MR analyses, along with additional sensitivity analyses.

Results: The association between CHCHD2 and an increased risk of PCOS was identified (OR=1.682; 95% CI=(1.231, 2.297); p=0.001). The discovery of CHCHD10 revealed a protective impact on the likelihood of PCOS (OR=0.828, 95% CI=0.698-0.981, p=0.029). The MR results were confirmed to be robust through the analysis of heterogeneity (p>0.05) and pleiotropy (p>0.05).

Conclusion: Our findings indicates that mitochondrial proteins CHCHD2 and CHCHD10 may play an important role in the pathogenesis of PCOS. Additional research is necessary to clarify the underlying mechanisms and investigate the potential of these proteins as targets for therapeutic intervention in PCOS.

A strong causal relationship has been established between two plasma mitochondrial complexes with coiled-coil-helix domains and polycystic ovary syndrome. The exact role of serum mitochondrial protein in polycystic ovary syndrome needs to be investigated *via* large-scale randomization trials or further studies.

under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited. **Keywords:** Polycystic ovary syndrome; Coiled-coil-helix-coiled-coil-helix domain Mitochondria; Genome wide association study; Mendelian randomization analysis

INTRODUCTION

Polycystic Ovary Syndrome (PCOS) is a prevalent hormonal condition that affects women during their childbearing years ^[1]. PCOS is characterized by hormonal imbalance, dysfunction of the ovaries and the presence of numerous cysts in the ovaries ^[2]. PCOS has been linked to various metabolic disorders, including insulin insensitivity, obesity and abnormal blood lipid levels ^[3]. The above-mentioned risk factors increase the likelihood of developing type 2 diabetes and cardiovascular disease ^[4,5]. The underlying mechanisms of PCOS remain unknown despite extensive research.

Mitochondria play an important role in cellular energy metabolism and physiological processes, including steroidogenesis, insulin signaling, oxidative stress response, apoptosis and mitochondrial dynamics ^[6]. Studies suggested a possible relationship between mitochondrial dysfunction and PCOS ^[7,8]. Mitochondrial dysfunction in women with PCOS may contribute to metabolic abnormalities, such as insulin resistance and obesity ^[9,10]. Impaired mitochondrial function in PCOS may decrease energy production and increase oxidative stress, contributing to insulin resistance and hormonal imbalances ^[11].

Numerous studies have shown that mitochondrial enzymes involved in energy production are diminished in women with PCOS. However, the precise mechanisms by which PCOS is associated with mitochondrial dysfunction remain poorly understood. Moreover, genetic and environmental factors such as obesity and insulin resistance are hypothesized to play a role in developing mitochondrial dysfunction in PCOS ^[12,13].

Recent studies have established that mitochondrial Coiled-Coil-Helix-Coiled-Coil-Helix Domain (CHCHD) proteins are involved in several cellular processes, such as mitochondrial function and response to oxidative stress ^[14,15]. However, the precise mitochondrial CHCHD proteins implicated in the development of PCOS and their causal connections to the disease are yet to be determined.

Understanding the cause-and-effect relationship between these mitochondrial proteins and PCOS could provide substantial insights into the pathogenesis of the disease and potentially reveal novel therapeutic targets. Mendelian Randomization (MR) is a statistical approach that uses genetic variants as Instrumental Variables (IVs) to investigate causal relationships between exposures and outcomes ^[16]. By minimizing biases from confounding and reverse causation, MR studies offer robust evidence ^[17] for causal associations.

The present investigation employed MR analysis to ascertain the causal relationship between mitochondrial CHCHD protein concentrations and PCOS risk. We employed genetic variants as IVs for plasma protein levels to evaluate their effect on the risk of PCOS. Furthermore, we examined the reverse causation by analyzing how PCOS affects the plasma levels of mitochondrial CHCHD proteins. This two-way method allowed us to understand the causal correlation between mitochondrial CHCHD proteins and PCOS. The findings of this study could have substantial implications for the identification, prevention and management of PCOS.

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Recognizing the causal link between mitochondrial CHCHD proteins and PCOS could aid the development of targeted therapies to restore mitochondrial function and improve PCOS-related metabolic abnormalities. Furthermore, determining the effect of PCOS on plasma concentrations of CHCHD proteins in mitochondria could help understand the possible role of mitochondrial dysfunction in the development of the syndrome.

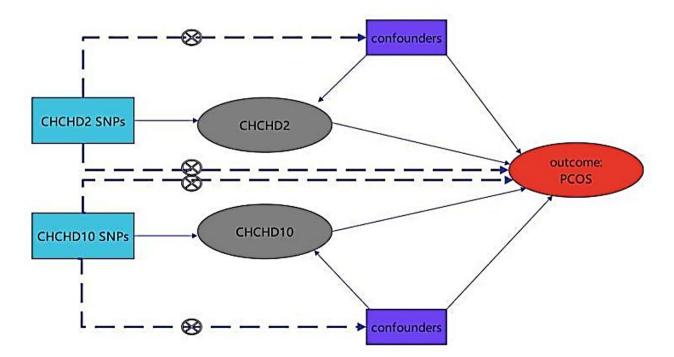
MATERIALS AND METHODS

Study design

To examine a causal relationship between PCOS and genetically predicted plasma levels of mitochondrial CHCHD proteins, we employed a two-sample MR strategy comprising two main stages. An MR analysis was initially performed using mitochondrial CHCHD proteins (CHCHD2 and CHCHD10) as exposures and PCOS as the outcome. In addition, an MR analysis was conducted in reverse, with PCOS as the independent variable and CHCHD2 and CHCHD10 as dependent variables. Notably, our study scrupulously maintains adherence to the following fundamental assumptions of MR analysis:

- The independent variables used in the analysis strongly correlate with exposure.
- The independent variables related to confounding factors have an insignificant influence on the relationship between exposure and the outcome.
- The independent variables solely affect the outcomes through their influence on exposure, excluding any other mechanisms. The study design is illustrated in Figure 1.

Figure 1. The flowchart of the MR study. It is an illustrative diagram of the two-way MR analysis. We probed causalities by selecting important IVs for CHCHD2, CHCHD10 and PCOS. The flowchart demonstrates the three fundamental assumptions of the MR analysis.



Data sources

GWAS summary data of CHCHD proteins and PCOS were obtained from the Integrative Epidemiology Unit (IEU) OpenGWAS database (https://gwas.mrcieu.ac.uk/). The plasma mitochondrial CHCHD proteins (CHDCHD2 and CHCHD10) were acquired from a comprehensive GWAS on 3301 European ancestry healthy adults (ID 'prot-a-534' and 'prot-a-535'). The genetic instruments for GWAS summary statistics of PCOS (642 cases and 118, 228 controls) were retrieved from the FinnGen Consortium (ID 'finn-b-E4_POCS').

Selection of the genetic instruments

Single Nucleotide Polymorphisms (SNPs) as IVs were utilized to assess the combined effect of mitochondrial CHCHD proteins on PCOS. We used p-values of 5×10^{-6} to select IVs for PCOS and plasma levels of mitochondrial CHCHD proteins. To ensure the selection of independent instruments, SNPs that did not overlap and were susceptible to LD clumping were chosen with a pairwise LD R² threshold below 0.001.

The genetic mutations were normalized by considering their impact while disregarding any palindromic mutations. The traits were analysed by employing available genetic summary statistics for the variants. Moreover, to maintain the integrity of the results, we abstained from any modification or manipulation of the IVs and the alleles linked to CHCHD2 and CHCHD10 were aligned in all the analyses. Instrument strength was determined by calculating the F-statistics and IVs with F-statistics greater than 10 were considered robust.

Statistical analysis

Inverse-Variance Weighted (IVW) was used as a key approach for univariate analysis in this study. A random-effects meta-analysis was performed to combine the results obtained from the individual SNPs. Additionally, weighted median, MR-Egger, weighted and simple mode analyses were conducted to ensure the causal direction was consistent. Cochran's test was performed in the IVW and MR-Egger models to assess heterogeneity among the included SNPs. Heterogeneity was considered significant if the p-value was <0.05.

Furthermore, we applied the MR-Egger regression to investigate potential pleiotropy, a method that allows for the simultaneous impact of genetic variants on multiple traits. In addition, MR-PRESSO detects and eliminates potential outliers that may introduce pleiotropy, consequently minimizing the probability of errors and bias due to pleiotropy. Statistical analyses were performed using R version 4.3.0.

RESULTS

Genetic IV selection for mitochondrial CHCHD proteins and PCOS

The current study used MR analysis to investigate the relationship between PCOS and CHCHD2 and CHCHD10. The research specifically examined the inverse correlation, with PCOS being identified as the risk and CHCHD proteins as the result.

Based on the selection criteria, 12 SNPs and 10 SNPs were found to be associated with CHCHD2 and CHD10, respectively. To explore the reverse causal effect, eight SNPs were chosen as credible IVs representing PCOS as the exposure. Supplementary Tables 1-3 contain detailed information on all the SNPs included.

The causal effect of CHCHD2 and CHCHD10 on PCOS

Table 1 displays three sensitivity analyses (IVW, weighted median and weighted mode) that confirmed the causal relationship between CHCHD10 and PCOS, whereas CHCHD10 and PCOS agreed with the two sensitivity analysis techniques (IVW and weighted median).

This study discovered a notable link between CHCHD2 and CHCHD10 and the likelihood of PCOS, with corresponding Odds Ratio (OR) values of 1.682 (95% CI=1.231-2.297; p=0.001) and 0.828 (95% CI=0.698-0.981; p=0.001), respectively. A visual representation of the estimated effects of each genetic mutation on the outcomes can be observed in the forest plot. Supplementary Figure 1 depicts a scatter plot depicting the estimated causal effects of each SNP on the risk of PCOS.

Exposure	Outcome	nSNP	Method	p-value		OR (95% CI)			
		12	MR Egger	0.151	⊢ ≎ →	1.656 (0.877 to 3.129)			
		12	Weighted median	0.004	⊢ ● →	1.809 (1.212 to 2.700)			
CHCHD2	PCOS	12	Inverse variance weighted	0.001	· · · ·	1.682 (1.231 to 2.297)			
		12	Simple mode	0.097	• • •	2.118 (0.941 to 4.767)			
		12	Weighted mode	0.025	↓ ————————————————————————————————————	2.266 (1.223 to 4.197)			
		10	MR Egger	0.252	HOH	0.834 (0.625 to 1.112)			
		10	Weighted median	0.034	Her	0.820 (0.683 to 0.985)			
CHCHD10	PCOS	10	Inverse variance weighted	0.029	Hei	0.828 (0.698 to 0.981)			
		10	Simple mode	0.443		0.804 (0.472 to 1.370)			
		10	Weighted mode	0.082	Harl	0.819 (0.670 to 1.000)			
0 0.5 1 1.5 2 2.5 3									
Note: MR=Mendelian Randomization; CI=Confidence Interval; OR=Odds Ratio; SNP=Single Nucleotide Polymorphism; CHCHD=Coiled-Coil-Helix-Coiled-Coil-Helix Domain-Containing Protein									

Table 1. Results of MR analysis between CHCHD proteins and PCOS.

Sensitive analysis

Heterogeneity was assessed using the Cochran Q-test, which yielded p-values for MR-Egger and IVW. No significant differences were observed between the selected SNPs in CHCHD2 and CHCHD10 (Table 2). No indications of directional pleiotropy were detected in the MR-Egger regression. Supplementary Figure 2 illustrates the results of the leave-one-out sensitivity analysis, revealing the causal connection between plasma CHCHD2, CHCHD10, and PCOS. The funnel plots of the associations between the SNPs in CHCHD2 and CHCHD10 and cancer risk were symmetrical, as presented in Supplementary Figure 3.

Table 2. Results of sensitivity analyses of CHCHD proteins on PCOS.

Exposure	Outcome		Cochran'	s Q test	Pleiotropy MR-Egger		p-value					
		MR-	Egger	IVW								
		Q	p-value	Q	p-value	Intercept	SE					
CHCHD2	COPD	14.096	0.0167	14.1	0.228	0.004	0.072	0.957				
CHCHD10	COPD	6.869	0.551	6.873	0.65	-0.001	0.004	0.994				
Note: MR=Mendelian randomization; IVW=Inverse-variance weighted; CHCHD=Coiled-coil-helix-coiled-coil-helix-coiled-coil-helix domain-containing protein; SE=Standard error of the beta												

DISCUSSION

This study examined the causal link between plasma twin mitochondrial proteins CHCHD2, CHCHD10 and PCOS using MR analysis. The MR analysis evaluates causality by employing genetic variants as IVs. By employing genetic variations that have been established to be associated with exposure (CHCHD2 and CHCHD10) but not with outcome (PCOS), it is possible to reduce the probability of confounding bias and reverse causation. Furthermore, two-sample MR analysis demonstrated a noteworthy causal connection between the plasma proteins of the CHCHD domain and PCOS. We discovered a correlation between genetically anticipated elevated levels of CHCHD2 and heightened

susceptibility to PCOS (OR=1.682, 95% CI=1.231-2.297, p=0.001). In contrast, an inverse relationship was noted, suggesting that genetically projected higher plasma concentrations of CHCHD2 were linked to a reduced risk of PCOS (OR=0.828, 95% CI=0.698-0.981, p=0.029). The current study found a potential causal association between CHCH2, CHCHD10 and PCOS development. Further research is required to elucidate the underlying mechanisms linking these proteins to PCOS.

Our findings exposed a notable and favorable connection between genetically anticipated plasma CHCHD2 concentrations and the risk of PCOS, while plasma CHCHD10 concentrations were negatively linked with PCOS risk. Mitochondrial respiration and oxidative phosphorylation are regulated by CHCHD2 ^[18]. Numerous neurodegenerative disorders, including Amyotrophic Lateral Sclerosis (ALS) and Parkinson's disease, have been associated with mutations in the *CHCHD2* gene ^[19,20]. Moreover, CHCHD10 is involved in regulating mitochondrial structure and function ^[21]. Frontotemporal dementia, a rare neurodegenerative disorder and cardiovascular diseases are linked to mutations in the *CHCHD10* gene ^[22,23]. *CHCHD2* and *CHCHD10* genes plays a vital role in maintaining the health and functionality of mitochondria, which is essential for cellular metabolism ^[24,25]. ATP, an energy substrate is synthesized *via* oxidative phosphorylation in mitochondria, earning them the moniker "powerhouses" of the cell ^[26]. Mitochondrial dysfunction can lead to various metabolic disorders, including mitochondrial diseases and neurodegenerative disorders ^[27].

Mitochondrial dysfunction may decrease ATP production and increase Reactive Oxygen Species (ROS) generation, harming cells and tissues ^[28,29]. Several studies reported that women with PCOS have impaired mitochondrial function in various tissues, including the ovaries, skeletal muscle and adipose tissues ^[30,32]. PCOS, insulin resistance and abnormal steroidogenesis have been linked to mitochondrial dysfunction ^[33]. Ovaries of the PCOS mouse model exhibited reduced mitochondrial DNA content and altered gene expression related to mitochondrial biogenesis and function, impairing follicle development and subsequently oocyte quality ^[34,35]. Additionally, mitochondrial dysfunction in PCOS may contribute to developing insulin resistance ^[36]. Mitochondrial dysfunction can delay the pathways responsible for insulin signaling, leading to reduced glucose uptake and increased ROS production ^[37,38]. Consequently, insulin resistance, a common characteristic of PCOS, may occur ^[39]. The precise mechanisms by which mitochondrial dysfunction is associated with PCOS remain unknown. Our findings further supported that the involvement of CHCHD2 and CHCHD10 in mitochondrial dysfunction plays a potential role in the development of PCOS. The interplay between genetic and environmental factors likely contributes to mitochondrial dysfunction in PCOS ^[40]. Disruptions of mitochondrial function may also be influenced by hormonal imbalances, including increased androgen and insulin levels ^[41,42].

The findings of this study could have substantial implications for understanding the underlying mechanisms of PCOS and identifying potential therapeutic targets. Targeting mitochondrial dysfunction and oxidative stress pathways may offer novel therapeutic and preventive strategies for PCOS. Further research is required to elucidate the distinct functions of CHCHD2 and CHCHD10 in PCOS development and explore possible interventions that can regulate their levels or influence their activity. Future research should focus on the specific molecular mechanisms underlying the association between CHCHD2 and CHCHD10 levels and metabolic abnormalities in PCOS.

CONCLUSION

In conclusion, this study indicates a possible causal link between twin mitochondrial proteins, CHCHD2 and CHCHD10, in the plasma and PCOS using MR analysis. Our results suggested that mitochondrial dysfunction may contribute to PCOS development. Additional investigations are required to clarify the fundamental processes and investigate the healing possibilities of focusing on the levels of CHCHD2 and CHCHD10 for PCOS management.

LIMITATIONS AND STRENGTHS

This study had some limitations. The MR analysis is predicated on the supposition that the genetic variants used as IVs are valid instruments and satisfy the requirements of instrumental variable assumptions. Despite utilizing previously validated genetic variants linked to CHCHD2 and CHCHD10 levels, the results may still be biased due to pleiotropy or horizontal pleiotropy. Notably, our research included individuals from a particular ethnic group, potentially restricting the applicability of our results to different demographics. As far as our knowledge goes, however, this is the first study to investigate the correlation between CHCHD proteins and PCOS. Future studies should verify these findings in diverse populations.

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CONFLICT OF INTEREST

The authors declare that they have no competing interests that might cause the work to be influenced by those interests in this manuscript.

ETHICS STATEMENT

As this was a re-analysis of published data, no ethical approval was required in this study.

REFERENCES

- 1. Zhuang C, et al. Cardiovascular risk according to body mass index in women of reproductive age with polycystic ovary syndrome: A systematic review and meta-analysis. Front Cardiovasc Med. 2022;9:822079.
- 2. Wang S, et al. Abnormal activation of tryptophan-kynurenine pathway in women with polycystic ovary syndrome. Front Endocrinol (Lausanne). 2022;13:877807.
- Guo F, et al. The lipid profiles in different characteristics of women with PCOS and the interaction between dyslipidemia and metabolic disorder states: A retrospective study in Chinese population. Front Endocrinol (Lausanne). 2022;13:892125.
- 4. Xiao B, et al. Inference of causal relationships between genetic risk factors for cardiometabolic phenotypes and female-specific health conditions. J Am Heart Assoc. 2023;12:e026561.

- 5. Duica F, et al. Impact of increased oxidative stress on cardiovascular diseases in women with polycystic ovary syndrome. Front Endocrinol (Lausanne). 2021;12:614679.
- 6. Post A, et al. Plasma creatine and incident type 2 diabetes in a general population-based cohort: The prevend study. Clin Endocrinol (Oxf). 2021;94:563-574.
- Dabravolski SA, et al. Mitochondrial dysfunction and chronic inflammation in polycystic ovary syndrome. Int J Mol Sci. 2021;22:3923.
- 8. Ruan Y, et al. CHCHD2 and CHCHD10 regulate mitochondrial dynamics and integrated stress response. Cell Death Dis. 2022;13:156.
- Rabah HM, et al. Novel insights into the synergistic effects of selenium nanoparticles and metformin treatment of letrozole-induced polycystic ovarian syndrome: Targeting PI3K/Akt signalling pathway, redox status and mitochondrial dysfunction in ovarian tissue. Redox Rep. 2023;28:2160569.
- Siemers KM, et al. Mitochondrial dysfunction in PCOS: Insights into reproductive organ pathophysiology. Int J Mol Sci. 2023;24:13123.
- 11. Moreno-Asso A, et al. Non-cell autonomous mechanisms control mitochondrial gene dysregulation in polycystic ovary syndrome. J Mol Endocrinol. 2021;68:63-76.
- Peng Y, et al. Novel mechanisms underlying anti-polycystic ovary like syndrome effects of electroacupuncture in rats: Suppressing SREBP1 to mitigate insulin resistance, mitochondrial dysfunction and oxidative stress. Biol Res. 2020;53:50.
- 13. Zhou Q, et al. Mitochondrial dysfunction caused by SIRT3 inhibition drives proinflammatory macrophage polarization in obesity. Obesity (Silver Spring). 2023;31:1050-1063.
- 14. Imai Y, et al. Twin CHCH proteins, CHCHD2 and CHCHD10: Key molecules of parkinson's disease, amyotrophic lateral sclerosis and frontotemporal dementia. Int J Mol Sci. 2019;20:908.
- 15. Wilson R, et al. Identification of key pro-survival proteins in isolated colonic goblet cells of winnie, a murine model of spontaneous colitis. Inflamm Bowel Dis. 2020;26:80-92.
- 16. Islam SN, et al. Reporting methodological issues of the mendelian randomization studies in health and medical research: A systematic review. BMC Med Res Methodol. 2022;22:21.
- 17. Katikireddi SV, et al. Assessing causal relationships using genetic proxies for exposures: An introduction to mendelian randomization. Addiction. 2018;113:764-774.
- 18. Purandare N, et al. The cellular stress proteins CHCHD10 and MNRR1 (CHCHD2): Partners in mitochondrial and nuclear function and dysfunction. J Biol Chem. 2018;293:6517-6529.
- 19. Trinh D, et al. The multi-faceted role of mitochondria in the pathology of parkinson's disease. J Neurochem. 2021;156:715-752.

- 20. Straub IR, et al. Loss of CHCHD10-CHCHD2 complexes required for respiration underlies the pathogenicity of a *CHCHD10* mutation in ALS. Hum Mol Genet. 2018;27:178-189.
- 21. Xia W, et al. CHCHD10 is dispensable for myogenesis but critical for adipose browning. Cell Regen. 2022;11:14.
- 22. Alici H, et al. Structures of the wild-type and S59L mutant CHCHD10 proteins important in amyotrophic lateral sclerosis-frontotemporal dementia. ACS Chem Neurosci. 2022;13:1273-1280.
- 23. Jiang T, et al. CHCHD2 and CHCHD10: Future therapeutic targets in cognitive disorder and motor neuron disorder. Front Neurosci. 2022;16:988265.
- 24. Liu W, et al. CHCHD2 regulates mitochondrial morphology by modulating the levels of Opa1. Cell Death Differ. 2020;27:2014-2029.
- 25. Ding M, et al. CHCHD10 modulates thermogenesis of adipocytes by regulating lipolysis. Diabetes. 2022;71:1862-1879.
- 26. Auger C, et al. Beyond mitochondria: Alternative energy-producing pathways from all strata of life. Metabolism. 2021;118:154733.
- 27. Cojocaru KA, et al. Mitochondrial dysfunction, oxidative stress and therapeutic strategies in diabetes, obesity and cardiovascular disease. Antioxidants (Basel). 2023;12:658.
- 28. Wei M, et al. Effect of fluoride on cytotoxicity involved in mitochondrial dysfunction: A review of mechanism. Front Vet Sci. 2022;9:850771.
- 29. Schaefer PM, et al. Combination of common mtDNA variants results in mitochondrial dysfunction and a connective tissue dysregulation. Proc Natl Acad Sci USA. 2022;119:e2212417119.
- 30. Zhang Q, et al. Mitochondrial and glucose metabolic dysfunctions in granulosa cells induce impaired oocytes of polycystic ovary syndrome through sirtuin 3. Free Radic Biol Med. 2022;187:1-16.
- 31. Yao Q, et al. Oxidative stress as a contributor to insulin resistance in the skeletal muscles of mice with polycystic ovary syndrome. Int J Mol Sci. 2022;23:11384.
- 32. Pruett JE, et al. Mitochondrial function and oxidative stress in white adipose tissue in a rat model of PCOS: Effect of SGLT2 inhibition. Biol Sex Differ. 2022;13:45.
- 33. Xia S, et al. Untargeted metabolomics analysis revealed lipometabolic disorders in perirenal adipose tissue of rabbits subject to a high-fat diet. Animals (Basel). 2021;11:2289.
- 34. Safaei Z, et al. Effect of vitamin D3 on mitochondrial biogenesis in granulosa cells derived from polycystic ovary syndrome. Int J Fertil Steril. 2020;14:143-149.
- 35. Reddy TV, et al. Polymorphisms in the TFAM and PGC1- α genes and their association with polycystic ovary syndrome among south Indian women. Gene. 2018;641:129-136.
- 36. Ding Y, et al. Mitochondria-targeted antioxidant therapy for an animal model of PCOS-IR. Int J Mol Med. 2019;43:316-324.

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- 37. Bourebaba N, et al. *Laurus nobilis* ethanolic extract attenuates hyperglycemia and hyperinsulinemia-induced insulin resistance in HepG2 cell line through the reduction of oxidative stress and improvement of mitochondrial biogenesis-possible implication in pharmacotherapy. Mitochondrion. 2021;59:190-213.
- 38. Tham YY, et al. Lauric acid alleviates insulin resistance by improving mitochondrial biogenesis in THP-1 macrophages. Mol Biol Rep. 2020;47:9595-9607.
- 39. Atef MM, et al. Therapeutic potential of sodium selenite in letrozole induced polycystic ovary syndrome rat model: Targeting mitochondrial approach (selenium in PCOS). Arch Biochem Biophys. 2019;671:245-254.
- 40. Szukiewicz D, et al. Modulation of the inflammatory response in Polycystic Ovary Syndrome (PCOS)-searching for epigenetic factors. Int J Mol Sci. 2022;23:14663.
- 41. Song L, et al. Androgen excess induced mitochondrial abnormality in ovarian granulosa cells in a rat model of polycystic ovary syndrome. Front Endocrinol (Lausanne). 2022;13:789008.
- 42. Kurdoglu Z, et al. Oxidative status and its relation with insulin resistance in young non-obese women with polycystic ovary syndrome. J Endocrinol Invest. 2012;35:317-321.