Application of CRISPR-Cas9 in Dairy Bacteriophage Research

Sidney Cobb*

Department of Food Science, University of Bordeaux, Bordeaux, France

Commentary

Received: 17-May-2024, Manuscript No. JFPDT-24-142153; Editor assigned: 21-May-2024, Pre OC No. JFPDT-24-142153 (PO); Reviewed: 4-June-2024, OC No. JFPDT-24-142153: Revised: 11-June-2024, Manuscript No. JFPDT-24-142153 (R); Published: 18-June-2024, DOI: 10.4172/2321-6204.12.2.005 *For Correspondence: Sidney Cobb, Department of Food Science, University of Bordeaux, Bordeaux, France E-mail: sidneycobb@iarc.fr Citation: Cobb S. Application of CRISPR-Cas9 in Dairy Bacteriophage Research. 2024; RRJ Food Dairy Technol. 12:005 Copyright: © 2024 Cobb S. This is an openaccess article distributed under the terms of the Creative Commons Attribution

License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABOUT THE STUDY

The advent of CRISPR-Cas9 technology has revolutionized genetic engineering, providing a powerful tool for precise genome editing. Initially discovered as a part of the bacterial immune system, CRISPR-Cas9 has found applications across various fields, including dairy bacteriophage research. Bacteriophages, viruses that infect bacteria, play a significant role in the dairy industry by influencing bacterial cultures used in the production of dairy products. This article delves into the application of CRISPR-Cas9 in dairy bacteriophage research, highlighting its potential to enhance dairy production, improve bacterial resistance, and ensure product safety.

CRISPR (Clustered Regularly Interspaced Short Palindromic Repeats) and Cas9 (CRISPR-associated protein 9) form a genetic system that allows bacteria to defend against invading viruses, including bacteriophages. When a bacteriophage infects a bacterium, the CRISPR system captures snippets of the viral DNA and integrates them into the bacterial genome. These serve as a genetic memory, enabling the bacterium to recognize and combat subsequent infections by the same virus.

Cas9, an endonuclease enzyme, is guided by RNA sequences derived from the CRISPR regions to target and cleave specific DNA sequences in the invading bacteriophage. This precise cutting mechanism has been harnessed in genetic engineering to edit genomes with unprecedented accuracy.

Research and Reviews: Journal of Food and Dairy Technology

One of the primary applications of CRISPR-Cas9 in dairy bacteriophage research is the enhancement of bacterial resistance to phage attacks. Lactic Acid Bacteria (LAB) are crucial in dairy fermentation processes, and bacteriophage infections can disrupt these processes, leading to product spoilage and economic losses. By employing CRISPR-Cas9, researchers can engineer LAB strains with enhanced resistance to specific bacteriophages. This is achieved by inserting synthetic CRISPR arrays that target and neutralize phage DNA, thereby protecting the bacterial cultures.

Phage therapy, the use of bacteriophages to target and eliminate pathogenic bacteria, is gaining traction as an alternative to antibiotics. In dairy production, CRISPR-Cas9 can be used to modify bacteriophages, making them more effective against harmful bacteria while sparing beneficial strains. This targeted approach ensures the maintenance of essential microbial communities in dairy fermentation, leading to improved product quality and safety.

CRISPR-Cas9 enables precise editing of bacteriophage genomes, allowing researchers to modify phages for specific applications. For instance, bacteriophages can be engineered to carry beneficial genes that enhance the metabolic activities of LAB, improving fermentation efficiency and product yield. Additionally, phages can be modified to include reporter genes that facilitate the detection and monitoring of bacterial infections in real-time.

Understanding the complex interactions between bacteriophages and their bacterial hosts is critical for developing effective phage-based interventions. CRISPR-Cas9 provides a powerful tool for dissecting these interactions at the genetic level. By selectively knocking out or modifying specific bacterial genes, researchers can identify key factors involved in phage resistance and susceptibility. This knowledge can guide the design of robust bacterial strains and phage therapies.

While the application of CRISPR-Cas9 in dairy bacteriophage research holds commitment, several challenges need to be addressed. The off-target effects of CRISPR-Cas9, where unintended genomic regions are edited, pose a risk to the stability and safety of engineered bacterial strains. Developing more precise and efficient CRISPR systems is crucial to overcoming this limitation. Moreover, the regulatory landscape for Genetically Modified Organisms (GMOs) varies across regions, affecting the adoption of CRISPR-engineered bacteria and phages in the dairy industry. Navigating these regulatory frameworks and ensuring consumer acceptance will be vital for the successful implementation of this technology.

The application of CRISPR-Cas9 in dairy bacteriophage research represents a significant advancement in the field of dairy technology. By enhancing bacterial resistance, facilitating phage therapy, enabling genomic editing, and elucidating phage-host interactions, CRISPR-Cas9 offers innovative solutions to longstanding challenges in dairy production. As research progresses and technology evolves, CRISPR-Cas9 has the potential to revolutionize the dairy industry, ensuring higher product quality, improved safety, and increased sustainability.