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Study of Antimicrobial Activity of Cynodon dactylon

Disha Sharma

Department of Biotechnology, Amity University, NODIA, India

Review Article

ABSTRACT

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*For Correspondence

Department of biotechnology, Amity University Nodia.

E-mail: dishasharma2112@gmail.com

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Plants are of most imperative wellsprings of medications since times indecent. Vast quantities of medications are being disconnected and separated from plants. The restorative plants are the wellsprings of optional metabolites and fundamental oils of remedial significance. The vital points of interest against the remedial utilization of therapeutic plants in different afflictions and scatters are their wellbeing other than practical, powerful and their simple accessibility. The utilization of and quest for medications and dietary supplements got from plants have quickened as of late. Abstract content goes here. Plants are of most imperative wellsprings of medications since times indecent. Vast quantities of medications are being disconnected and separated from plants. The restorative plants are the wellsprings of optional metabolites and fundamental oils of remedial significance. The vital points of interest against the remedial utilization of therapeutic plants in different afflictions and scatters are their wellbeing other than practical. powerful and their simple accessibility. The utilization of and quest for medications and dietary supplements got from plants have quickened as of late.

INTRODUCTION

Large numbers of drugs are being isolated and extracted from plants. The medicinal plants are the sources of secondary metabolites and essential oils of therapeutic importance. The important advantages against the therapeutic use of medicinal plants in various ailments and disorders are their safety besides economical, effective and their easy availability ^[1-3]. Cynodon dactylon is a perennial grass belonging to family Poaceae that has a variety of medicinal properties. It is cultivated throughout the tropics and subtropics. Whole herb and its root stalk are used for medicinal use ^[4-6]. It is native to north and east Africa, Asia and Australia and southern Europe. In Ayurveda Cynodon dactylon shows many pharmacological activities like antidiabetic, antioxidant, antidiarrheal, hepatoprotective, antiulcer, immunomodulator, CNS depressant, antimicrobial and germicidal ^[7-10]. An investigation showed that the aqueous extractof Cynodon dactylon has high antidiabetic potential along with significant hypoglycemic and hypolipidemic effects. The plant contains crude proteins, carbohydrates, and mineral constituents, oxides of magnesium, phosphorous, calcium, sodium and potassium ^[11-13].

The whole plant affords β-sitosterol, flavonoids, alkaloids, glycosides and triterpenoids. Cynodon dactylon contain many chemical constituents like Hexadecanoic acid, Linolenic acid, ethyl ester, Hydroquinone, d-mannose ^[14-16]. Synonyms: Agrostis bermudiana Tussac; Agrostis filiformis J.Koenig; Agrostis linearis Retz; Agrostis stellata Willd; Capriola dactylon L. Kuntze; Capriola dactylon L. Hitchc; Chloris cynodon Trin. Nom. illeg; Chloris maritima Trin ^[17,18]. Common Name: Bermuda Grass; Assamese: Dubari; Bengali: Durba; Hindi: Doob, Dobri; Kannada: garikehullu, balli garike, ambate hullu; Manipuri: Tingthou; Marathi: haryali, dhurva; Mizo: phaitualhnim; Oriya: dubbo ghas; Sanskrit: Niladurva, saddala, ananta; Tamil: Arugampillu, Arukampillu, muyalpul; Telugu: Ghericha, gerichagaddi; Urdu: Doob. Common Name: Bermuda Grass; Assamese: Dubari; Bengali: Durba; Hindi: Doob, Dobri; Kannada: garikehullu, balli garike, ambate hullu; Manipuri: Tingthou; Marathi: haryali, dhurva; Mizo: phaitualhnim; Oriya: dubbo ghas; Sanskrit: Niladurva, saddala, ananta; Tamil: Arugampillu, Arukampillu, muyalpul; Telugu: Ghericha, gerichagaddi; Urdu: Doob. Common Name: Bermuda Grass; Assamese: Dubari; Bengali: Durba; Hindi: Doob, Dobri; Kannada: garikehullu, balli garike, ambate hullu; Manipuri: Tingthou; Marathi: haryali, dhurva; Mizo: phaitualhnim; Oriya: dubbo ghas; Sanskrit: Niladurva, saddala, ananta; Tamil: Arugampillu, Arukampillu, muyalpul; Telugu: Ghericha, gerichagaddi; Urdu: Doob ^[19-22].

CHEMICAL CONSTITUENTS

It contains crude proteins, carbohydrates and minerals constituents oxides of magnesium phosphorous, calcium, sodium and potassium. The whole plant affords sitosterol and carotene and potassium. Other compounds like vitamin c, cartone, palmonic acid, triterpenoids, alkaloids ergonovine and ergonovinine etc are also present ^{[25].}

Used parts of cynadon dactylon & preparation

Stem and leaves; Prepared form for study and use, the leaves are dried in sunlight for complete moisture evaporation. Un-required parts of the plants are separated and discarded while the rest of the part is grinded to obtain the powdered form for various research experiments use ^{[26].}

Selection of culturing media

The selection of media is an important part for antimicrobial and microbiological experimentation and study. Culturing media are the compositions of various macro and micro-molecules that provides and facilitates the growth of microorganism in in-vitro condition ^[27-29]. In today's industrialization period many companies are producing such ready to use media for experimentation. Variety of media depends on the choice of stain of microorganism. Different microorganism requires different growth requirement thus choice of media depends on the selection of strain that has be studied ^[30,31].

Antimicrobial activity

The four main mechanisms by which microorganisms exhibit resistance to antimicrobials are: Drug inactivation or modification; for example, enzymatic deactivation of penicillin G in some penicillin-resistant bacteria through the production of β -lactamases [32-36]. Most commonly, the protective enzymes produced by the bacterial cell will add an acetyl or phosphate group to a specific site on the antibiotic, which will reduce its ability to bind to the bacterial ribosomes and disrupt protein synthesis; Alteration of target site: for example, alteration of PBP-the binding target site of penicillin's-in MRSA and other penicillin-resistant bacteria [37,38]. Another protective mechanism found among bacterial species is ribosomal protection proteins [39]. These proteins protect the bacterial cell from antibiotics that target the cell's ribosomes to inhibit protein synthesis. The mechanism involves the binding of the ribosomal protection proteins to the ribosomes of the bacterial cell, which in turn changes its conformational shape [40-42]. This allows the ribosomes to continue synthesizing proteins essential to the cell while preventing antibiotics from binding to the ribosome to inhibit protein synthesis; Alteration of metabolic pathway: for example, some sulfonamide-resistant bacteria do not require Para-amino benzoic acid PABA, an important precursor for the synthesis of folic acid and nucleic acids in bacteria inhibited by sulfonamides, instead, like mammalian cells, they turn to using preformed folic acid; Reduced drug accumulation: by decreasing drug permeability or increasing active efflux pumping out of the drugs across the cell surface [43-47]. These specialized pumps can be found within the cellular membrane of certain bacterial species and are used to pump antibiotics out of the cell before they are able to do any damage. These efflux pumps are often activated by a specific substrate associated with an antibiotic [48-50].

Preparation of Plant Extracts

The thimble filled with about 55 g of each plant sample Stem, Root and Flower powder was subjected to various solvents of increasing polarity for successive use ^[51,52]. The solvents were evaporated to dryness in the rotary evaporator to obtain the solvent free extracts. The plant extracts so obtained were lyophilized at a definite temperature. The extracts powder was stored in airtight bottles at 37 °C till ^[53,54].

Disc Diffusion Method

For antimicrobial study the most commonly used method for anaylsis is the disc diffusion method. Kirby-Bauer antibiotic testing is a strategy to decide the affectability of microorganisms to particular antimicrobial medications; more noteworthy medication adequacy yields bigger organism free zones encompassing medication containing circles after overnight development on strong media ^[55,56].

Zone Of Inhibition

This is a territory of media where microscopic organisms can't develop, because of nearness of a medication that obstructs their development ^[57].

DISCUSSION

From the various reaches results cyandon dactylon has comes up with promising result. when test with various microorganism strains of both gram positive and gram negative bacterials like Bacillus subtilis BS, Bacillus atrophaeus BA and Staphylococcus aureus S, Escherichia coli EC, Pseudomonas aeruginosaPA, Salmoneella typhi ST, Kleibsiella pneumonia KP, Erwiniacarotovora Agrobacterium tumifaciens EC and AT and a fungal strain, Candida albicans CA and many more other strain ^[58-63]. The antimicrobial action of the leaves of cynadon dactylon has been found by different solvent extracts including solvents like water, ethanol, alcohol as well as crude extract ^[64-69].since the use of these solvents as extracting agents has been proved to be more efficient in extorting the active compounds ^[70-74]. All the extracts exhibited different degree of inhibition against selected gram positive and gram negative bacteria due to their varying polarity and solubility ^[75]. The polarity of the antibacterial compounds makes

them more readily extracted by organic solvents without negatively affecting the bio activity against bacterial species [76-80].

CONCLUSION

The medicinal plants are the sources of secondary metabolites and essential oils of therapeutic importance. The grass and cultivation is not an essential, therefore the species may easily be explored for human benefits from natural habitat. The present researches provide and proof with an evidence that the cyanndon dactylon has proven as beneficial gift to mankind. Intense antimicrobial potential of cynadon dactylon leaves extracts against gram negative bacteria, gram positive and a fungus. The findings suggest that the plant could be regarded as promising alternative for development of efficient and effective drug from natural source that can be used for therapy of infectious diseases. The most active extracts could be subjected for further pharmacological evaluation by isolation of the therapeutic antimicrobials and further research on this plant can specify its pharmaceutical application.

REFERENCES

- 1. Hassan W, et al. Antimicrobial Activity of Cinnamomum tamala Leaves. J Nutr Disorders Ther 2006; 6: 6.
- 2. Mir MA, et al. Antimicrobial Activity of Various Extracts of Taraxacum officinale. J Microb Biochem Technol 2006; 8:20-25.
- 3. Mateen Ayesha, et al. A study of antimicrobial activity of few medicinally important herbal single drugs extracted in ethanol, methanol and aqueous solvents. Nat Prod Chem Res 2005; 3:6-17.
- 4. Mir MA, et al. Antimicrobial Activity of Various Extracts of Taraxacum officinale. J Microb Biochem Technol 2005; 25:8-20.
- 5. Lakehal S, et al. Essential Oil Composition and Antimicrobial Activity of Artemisia herba–alba Asso Grown in Algeria. Med chem 2006; 6:435-439.
- 6. Krishnaveni V. Analysis of Chemical Components and Antimicrobial Activity on Vetiver Extract for Home Textile Applications. J Textile Sci Eng 2006; 7: 6-259.
- 7. Amer HH, et al. Synthesis and Antimicrobial Activity of New Synthesized Benzimidazole Derivatives and their Acyclic Nucleoside Analogues. Organic Chem Curr Res 2006; 5: 54-57.
- 8. Prodan AM, et al. Antimicrobial activity of collagen/silver doped hydroxyapatite composites against grampozitive and gram-negative bacteria. Chem Sci J 2016; 4: 51-57.
- Chintaluria AK and Komarrajub AL. Antimicrobial activity of essential oils from selected culinary herbs of Zingiberaceae and detection of antimicrobial compounds using TLC- Bioautography. Nat Prod Chem Res 2005; 5: 59-67.
- 10. Aliyu M Musa, et al. A novel C-prenylated benzo-lactone possessing antimicrobial activity from the rhizome of Cissus cornifolia. Clin Pharmacol Biopharm2015; 3:9-18.
- 11. Amin Mir M, et al. Antimicrobial Activity of Various Extracts of Taraxacum officinale. J Microb Biochem Technol 2016; 8:210-215.
- 12. Lakehal S, et al. Essential Oil Composition and Antimicrobial Activity of Artemisia herba–alba Asso Grown in Algeria. Med chem Los Angeles 2016; 6:435-439.
- 13. Krishnaveni V. Analysis of Chemical Components and Antimicrobial Activity on Vetiver Extract for Home Textile Applications. J Textile Sci Eng 2016; 6:240-259.
- 14. Amer HH, et al. Synthesis and Antimicrobial Activity of New Synthesized Benzimidazole Derivatives and their Acyclic Nucleoside Analogues. Organic Chem Curr Res 2016; 5: 145-157.
- 15. Hassan W, Kazmi SNZ, Noreen H, Riaz A, Zaman B. Antimicrobial Activity of Cinnamomum tamala Leaves. J Nutr Disorders Ther 2016; 201; 6: 185-190.
- 16. Kumar S, et al. Preliminary Phytochemical Screening and Antimicrobial Activity of Salvadora persica Linn. Extracts against Oral Pathogens. Fungal Genom Biol 2016; 6: 145-131
- 17. Xu L, et al. Antimicrobial Activity and Identification of Actinomycete Strains from a Folk Medicinal Soil in China. Appli Micro Open Access 2015; 1: 98-105.
- 18. Ale CE, et al. Microencapsulation of Bovine Vaginal Lactobacilli in Alginate Using Emulsiongelation: Freezedrying, Storage and Antimicrobial Activity. J Bioprocess Biotech 2015; 5: 255-262.
- 19. Singh BR, et al. Antimicrobial Activity of Citronella Essential Oil on Antimicrobial Drug Resistant Bacteria from Veterinary Clinical Cases. Clin Med Biochemistry Open Access 2015; 1: 93-106.
- 20. Lee JH, Choi HJ. Antiviral and Antimicrobial Activity of Medicinal Plant Extracts. J Microb Biochem Technol 2015; 7: 288-231.
- 21. Figueiras A, et al. Preparation and characterization of Trimethoprim inclusion complex with Methyl-β-Cyclodextrin and determination of its antimicrobial activity. Pharm Anal Acta 2015; 6: 401-405

- 22. Sharma MC. An Insight into Antimicrobial Activity Substituted Benzimidazole Derivatives through QSAR Studies. Drug Des 2015; 4: 122-128.
- 23. Rani VE and Kumar KD. Synthesis and Antimicrobial Activity of Some Novel N-Mannich Bases of Imidazole Phenylazetidin-2-one. Med chem 2015; 5: 156-159.
- 24. Chairman M, et al. Phytochemical Screening and Antimicrobial Activity of Curcuma longa Natural Dye. Gen Med Los Angel2015; 3: 169-171.
- 25. Hemanta MR, et al. Analysis of Traditional Food Additive Kolakhar for its Physico-Chemical Parameters and Antimicrobial Activity. J Food Process Technol 2014; 5: 383-387.
- 26. Singh K, et al. Evaluation of Antimicrobial Activity of Synthesized Silver Nanoparticles usingPhyllanthus amarus and Tinospora cordifolia Medicinal Plants. J Nanomed Nanotechnol 2015; 5: 249-250.
- 27. Deepa R, et al. Evaluation of Antimicrobial Activity and Antioxidant Activity by Electrochemical Method of Ethanolic Extract of Pterocarpus marsupium Roxb Bark. J Biotechnol Biomater 2014; 4: 166-169.
- 28. Vasconcelos WA, et al. Bioactive Glass-Green and Red Propolis Association: Antimicrobial Activity Against Oral Pathogen Bacteria. Nat Prod Chem Res 2014; 2: 151-154
- 29. Alwan SM, et al. Synthesis and Preliminary Antimicrobial Activity of New Schiff Bases of Pyrido [1,2-a] Pyrimidine Derivatives with Certain Amino Acids. Med chem 2014; 4: 687-639.
- 30. Gortzi O, et al. Study of Antioxidant and Antimicrobial Activity of Chios Mastic Gum Fractions Neutral, Acidic Before and After Encapsulation in Liposomes. J Food Process Technol 2014; Volume: 5-355
- 31. Kerru N et al. Novel Thienopyrimidine Derivatives Containing 1,2,4-triazoles and 1,3,4-oxadiazoles as Potent Antimicrobial Activity. Med chem 2014; 4: 624-629.
- 32. Abou-Dobara MI, et al. Thermal Properties, Antimicrobial Activity of Azo Complexes and Ultrastructure Study of Some Affected Bacteria. J Microbial Biochem Technol S2014; 3: 6-12.
- 33. Sharma G, et al. Synthesis, Characterization and Antimicrobial Activity of Abelia grandiflora Assisted AgNPs. J Microb Biochem Technol 2014; 6: 278
- 34. Shahaby OE, et al. Evaluation of Antimicrobial Activity of Water Infusion Plant-Mediated Silver Nanoparticles. J Nanomed Nanotechol 2013; 4: 175-178.
- 35. Zalabani SM et al. Anti-inflammatory and Antimicrobial Activity of the Different Conyza dioscoridis L. Desf. Organs. Biosafety 2013; 2: 106-107.
- 36. Khan SA, et al. Impaired Histatin-5 Levels and Salivary Antimicrobial Activity against C. albicans in HIV Infected Individuals. J AIDS Clin Res 2013; 4: 191-193.
- 37. Ahmad J and khan I. Evaluation of Antioxidant and Antimicrobial Activity of Ficus Carica Leaves: an In Vitro Approach. J Plant Pathol Microb 2013; 4: 154-157
- 38. Hasan HA, et al. Chemical Composition and Antimicrobial Activity of the Crude Extracts Isolated from Zingiber Officinale by Different Solvents. Pharmaceut Anal Acta 2012; 3: 181-184
- 39. Bueno J. In Vitro Antimicrobial Activity of Natural Products Using Minimum Inhibitory Concentrations, Looking for New Chemical Entities or Predicting Clinical Response. Med Aromat Plants 2012; 1: 112-113.
- 40. Lingham T, et al. Antimicrobial Activity of Vinegar on Bacterial Species Isolated from Retail and Local Channel Catfishlctalurus punctatus. J Food Process Technol S2012; 11: 1-16.
- 41. Anehosur GV, et al. Synthesis and Determination of Antimicrobial Activity of Visible Light Activated TiO2 Nanoparticles with Polymethyl Methacrylate Denture Base Resin Against Staphylococcus Aureus. J Gerontol Geriatric Res 2012; 1: 104-103.
- 42. Nicolete R and Nicolete LDF Microencapsulated Leukotrienes Augment Antimicrobial Activity against Infections. J Cell Sci Ther S 2012; S5-001.
- 43. Santos RJ Jr, et al. Antimicrobial Activity of Broth Fermented with Kombucha Colonies. J Microbial Biochem Technol 2009; 2: 1065-1078.
- 44. Batool Z, et al. Antimicrobial Resistance of ESBL Producing Coliforms Isolated from Retail Meat Samples. J Biom Biostat 2016; 7: 315-316.
- 45. Anuradha S De et al .Antimicrobial Resistance among Commonly Encountered Bacteria Isolated in 2013 The ESKAPE Menace. Intern Med 2015; 5: 194-195.
- 46. Ravalli R, et al. A Review on Antimicrobial Resistance in Developing Countries. Biochem Pharmacol 2015; 4: r001.
- 47. Adekunle OC and Onilude AA .Antimicrobial Resistance and Plasmid Profiles of Campylobacter Species from Infants Presenting with Diarrhoea in Osun State, Nigeria. J Med Microb Diagn 2015; 4: 171-172.

- 48. Guney AK, et al. A Study on Class I Integrons and Antimicrobial Resistance among Clinical Staphylococci Isolates from a Turkish Hospital. Clin Microbial 2014; 3: 169-173.
- 49. Edward PC Lai .Biochemical Challenges in Combating Antimicrobial Resistance. Biochem Anal Biochem 2014; 3:e150.
- 50. Garrido AM et al. Antimicrobial Resistance in Enterococci. J Infect Dis Ther 2014; 2: 150-151.
- 51. Mahero M, et al. Antimicrobial Resistance and Presence of Class 1 Integrons in Salmonella Serovars Isolated from Clinical Cases of Animals and Humans in North Dakota and Uganda. Clin Microbial 2013; 2: 127-128.
- 52. Chaudhary M and Payasi A. Rising Antimicrobial Resistance of Pseudomonas aeruginosa Isolated from Clinical Specimens in India. J Proteomics Bioinform 2013; 6:009.
- 53. Dias C, et al. Antimicrobial Resistance Patterns of Aeromonas spp. Isolated from Ornamental Fish. J Aquacult Res Dev 2012; 3: 130-131.
- 54. Karlton-Senaye B, et al. Interaction of Gums and Antimicrobial Agents on Susceptibility of SelectedFoodborne Pathogens. J Antimicro 2016; 2: 120-121.
- 55. El-Attar MAZ, et al. Design and Synthesis of Some New 1,2,4-Triazolo[4,3-a]Quinoxaline Derivatives as Potential Antimicrobial Agents. Med chem 2015; 5: 494-495.
- 56. Sharma MC. 2D QSAR Studies on a Series of Benzimidazole Type of Fluconazole Analogues as Antimicrobial Agents. J Nanomedine Biotherapeutic Discov 2012; 5: 131-133.
- Dong X, et al. Superior Antibacterial Activity of Photochemical Synthesized Ag-CNT Composites and their Synergistic Effects in Combination with other Antimicrobial Agents. J Nanomed Nanotechnol 2015; 6: 278-279.
- 58. Yan L, et al. In Vitro Synergism Testing Of Three Antimicrobial Agents against Multidrug-Resistant and Extensively Drug- Resistant Mycobacterium Tuberculosis by Checkerboard Method. J Mol Pharm Org Process Res; 2015; 2: 123.
- 59. Raghunath BT, et al. Synthe sis and Antimicrobial Activity of Benzo[H][1,6]Naphthyridine Derivatives. J Org Inorg Chem 2016; 2: 1.
- 60. Eryilmaz M. New Generation Disinfectants. J Pharm Microbiol 2016; 2: 1.
- 61. Aseer Manilal, et al. An In Vitro Antibacterial and Cytotoxic Potentials of Bioactive Metabolites Extracted from Padina tetrastromatica. Transl Biomed 2016; 7: 1.
- 62. Ly SY, et al. The In-Vivo Diagnostic Assay of Antimicrobial Activity Using Modified Probe. Int J Drug Dev & Res 2015; 7: 059.
- 63. Shanmugapriya K, et al. Evaluation of antioxidant, anti-inflammatory and antimicrobial activities of two different extracts of Camellia sinensis L. O. Kuntze. Int. J. Drug Dev. & Res 2013; 5: 3-6.
- 64. K. Mohana Priya and Samanta S. Khora Antimicrobial, Hemolytic and Cytotoxic activities of the Puffer Fish
- 65. Arothron hispidus from the Southeast Coast of India.Int. J. Drug Dev. & Res 2013; 5: 321-322.
- 66. Taneesha Chawla, et al. Antimicrobial activity of spices like cloves Cardamom and Cinnamon on Bacillus and Pseudomonas 2014; 6: 3-7.
- 67. Kapoor Nisha, et al. GC-MSAnalysis and anti-microbial activity of PsidiumGuajava leaves grown in Malva region of India,Int. J. Drug Dev. & Res 2011; 3: 245
- Pundir Ram Kumar and Bishnoi Shreya. Antimicrobial activity of Mitragyna parvifolia barks and Butea monosperma leaves extracts against human pathogenic microbial strains. Int. J. Drug Dev. & Res2011; 3: 147
- 69. Sulbha Lalsare, et al. Anti-Inflammatory and Antimicrobial activity of Flacourtia Ramontchi Leaves. Int J. Drug Dev. & Res 2011; 3: 313
- 70. Manish Gupta et al. Synthesis and Antimicrobial evaluation of 3,5-pyrazolidine-Dione substituted 4-Quinolone derivatives. Int J Drug Dev & Res 2011; 3: 239
- 71. Nagaraja TS, et al. Evaluation of Antimicrobial activity of Erythrina mysorensis gamb. Int J. Drug Dev. & Res 2011; 3: 202
- 72. Al-Manhel AJ, et al. Effect of Aqueous and Alcoholic Plant Extracts on Inhibition of Some Types of Microbes and Causing Spoilage of Food. J Nutr Food Sci 2015; 5: 006.
- 73. Antibacterial, Cytotoxic and Antioxidant potential of Methanolic extract of Phyllanthus Acidus L", Int J. Drug Dev. & Res 2011; 3: 161.
- 74. Pattari Lohitha, et al. Invitro Evaluation Of Antimicrobial Activity Of Butea Monosperma L Leaf Hexane: Ethanol [1:1ratio] Extract, Int. J. Drug Dev 2011; 3: 272
- 75. Abu Osman, et al. Antimicrobial Investigation on Manilkara zapota L. P. Royen? Int. J. Drug Dev. & Res 2011; 3: 190.
- 76. Hiba Hazim and Hamid Al-Yousuf. Antibacterial activity of Ficus carica L. extract against six bacterial strains. Int J Drug Dev. & Res 2012; 4: 310.
- 77. Gautam Keerti and Kumar Padma. Evaluation of Phytochemical and Antimicrobial study of Extracts of Vitex negundo Linn. Int. J. Drug Dev. & Res 2012; 4: 199.

- 78. Karumanchi Anupama Devi, et al. Synthesis and Antimicrobial activity of some Quinazolinones Derivatives.Int. J. Drug Dev. & Res 2012; 4: 327.
- 79. Prabhsimran Singh Sandhu, et al. Screening of Antimicrobial activity of Aqueous extracts of Leaves, Flower and Stem of Eclipta alba. Int. J. Drug Dev. & Res 2012; 4: 147.
- 80. P. Malairajana, et al. Preliminary Antimicrobial Screening Of SomeIndian Medicinal Plants Part I.Int. J. Drug Dev. & Res 2012; 4: 137.